

Synthesis and characterization of sugar-based methacrylates and their random copolymers by ATRP

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Abstract. Various sugar-based methacrylate monomers have been prepared and randomly copolymerized with methyl methacrylate (MMA) using classical atom transfer radical polymerization (ATRP). Firstly, four different sugar-based methacrylates are synthesized by two-step method: (i) etherification of protected monosaccharides with epichlorohydrin and (ii) following ring-opening reaction of obtained epoxides with methacrylic acid (MAA) in the presence of triethylamine. Next, these monomers are copolymerized with MMA via ATRP at 90 °C to obtain corresponding random copolymers. The molecular weights of the copolymers are determined by both GPC (gel permeation chromatography) and ¹H-NMR (nuclear magnetic resonance spectroscopy) analyses and found as $10600\sim16800$ and $12200\sim18500$ g/mol, respectively. Moreover, the copolymer compositions are also determined by ¹H-NMR analysis using characteristic signals of the monomers and found as about $94.1\sim97.8\%$, which are good agreement with feeding ratio. In addition, the glass transition temperatures of copolymers are found as $101.2\sim102.9$ °C by changing type and composition of sugar-based methacrylate monomers. Overall, a series of well-defined random copolymers comprising different sugar-based methacrylates and methyl methacrylates were successfully synthesized by classical ATRP method.

Keywords: polymer synthesis, atom transfer radical polymerization, random copolymers, sugar based monomers

1. Introduction

Nearly all commercial polymers are produced from petroleum sources, but they may come from renewable resources in the future. Glycopolymers are synthetic polymers obtained from natural sugar based monomers and possess structural diversity, multiple functionalities and innocuousness for human health [1–4]. Because of their fascinating properties such as biocompatibility, low prices and readily available precursors, they have been utilized in medicine and biotechnology applications [5]. The commercial availability of a variety of carbohydrates provides access to a wide range of glycomonomer derivatives. The most common synthetic approaches are chemical connection of a vinyl group with a suitably protected

carbohydrate, glycosylation of a halogenated sugar, Grignard reaction and enzymatic transesterification reactions [6]. Polymerization of these monomers can usually be achieved by free radical, controlled radical, anionic, cationic, ring-opening or ring-opening metathesis polymerization using monomers having pendant sugar units (glycomonomers) [7]. Among them, atom transfer radical polymerization (ATRP) permits the synthesis of well-defined (co)polymers from a wide selection of monomers and initiators under mild conditions [8–23]. To date, a number of well-defined glycopolymers from unprotected monomers have been successfully prepared by ATRP [24–28] or other controlled radical polymerization methods such as nitroxide mediated radical polymerization

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(NMP) [29, 30], reversible addition-fragmentation chain transfer (RAFT) [31–34] and cyanoxyl-mediated radical polymerization [35, 36].

In the present study, a series of well-defined random copolymers comprising different sugar-based methacrylates and methyl methacrylates were synthesized by ATRP and their structures were investigated by spectroscopic, chromatographic and thermal analyses. For this purpose, four different sugarbased methacrylates were firstly synthesized by twostep method: (i) etherification of protected monosaccharides with epichlorohydrin and (ii) subsequent ring-opening reaction of the products with methacrylic acid. The successful ATRP of these monomers in the presence of methyl methacrylate enabled to prepare well-defined random copolymers. Finally, the structure of the resulting copolymers was verified by ¹H-NMR spectroscopy, gel permeation chromatography and differential scanning calorimeter analyses.

2. Experimental Part2.1. Materials

D-Glucose, D-galactose and D-mannose were purchased from Merck EMD Millipore Corporation (Darmstadt, Germany) and used as received. Trichloroacetaldehyde (TCAA) was obtained by freshly distillation of chloral hydrate (≥98%, Sigma-Aldrich Chemical Corporation, Steinheim, Germany) treating with sulphuric acid under inert atmosphere. Epichlorohydrin (ECH, ≥99.0%), triethylamine (TEA, ≥99%), tetrabutylammonium bromide (TBAB, ≥98%), methacrylic acid (MAA, ≥99 %) were purchased from Merck EMD Millipore Corporation (Darmstadt, Germany). Dimethylformamide (DMF, ≥99.8%, Sigma-Aldrich Chemical Corporation, Steinheim, Germany) was dried on 4A molecular sieve. Protected furanosidic sugars D-glufrn, D-manfrn, SD-glufrn, D-galfrn were prepared according to literature respectively [37–40]. TLC (thin layer chromatography) and column chromatography were performed on precoated aluminum plates (Merck 5554) and silicagel G-60 (Merck 9385) respectively. Hexane-ethyl acetate (7:3) was used for TLC and column chromatography. Methyl methacrylate (MMA, 99%, Sigma-Aldrich Chemical Corporation, Steinheim, Germany) was passed through a basic alumina column to remove the inhibitor and stored in the freezer under nitrogen prior to use. N,N,N',N",N"-Pentamethyldiethylenetriamine (PMDETA; 99%, Sigma-Aldrich Chemical Corporation, Steinheim, Germany) was used as ligand and distilled before to use. The metal catalyst, copper(I) chloride (CuCl, 99.99%,Sigma-Aldrich Chemical Corporation, Steinheim, Germany) and the initiator, ethyl 2-bromopropionate (EtBrP, 99%, Sigma-Aldrich Chemical Corporation, Steinheim, Germany) were used without any purification. All solvents were purified by conventional distillation and drying procedures.

2.2. Instrumentation

Intermediates of the ¹H-NMR (Palo Alto, California, USA) analyses were recorded by a Varian 400 MHz NMR spectrometer at room temperature in CDCl₃ with tetramethylsilane as internal standard and chemical shifts were reported in ppm. The Perkin-Elmer (Waltham, USA) FT-IR Spectrum Two Spectrometer equipped with a diamond ATR device was used for Fourier transform infrared (FT-IR) analysis.

Gel permeation chromatography analyses were carried using a Viscotek GPCmax consisting of a pump module (GPCmax, Viscotek, Houston, TX) with flow rate 1 mL/min, a combined light-scattering (Model 270 dual detector, Viscotek), and a refractive index (RI) detector (VE 3580, Viscotek). Injections were done by an auto-sampler system, a 50 µL injection volume was used. The RI detector was calibrated using narrow molecular weight polystyrene standards. The light-scattering detector ($\lambda_0 = 670 \text{ nm}$) included two scattering angles: 90 and 7. Two columns (LT5000L, Mixed, Medium Organic 300×8 mm and LT3000L, Mixed, Ultra-Low Organic 300×8 mm) with a guard column (TGuard, Organic Guard Column 10×4.6 mm) were used for the tetrahydrofuran eluent at 35 °C. The data were analyzed using Viscotek OmniSEC Omni-01 software. Differential scanning calorimetry (DSC) was performed on a Perkin-Elmer (Waltham, USA) Diamond equipment under nitrogen flow (10 mL/min.) with a heating rate of 10°C/min.

2.3. General procedure for etherification reactions of sugars with epichlorohydrin

Epichlorohydrin (4 mL, 51 mmol), aq. NaOH (50%, 10 mL) and TBAB (0.5 g, 1.6 mmol) were stirred for 30 min at r.t. Then corresponding protected sugars (**D-glufrn**, **D-manfrn**, **SD-glufrn** and **D-galfrn**) (2 g, 5.8 mmol) were added slowly to this mixture at approximately 5 °C and the reaction continued at the same temperature for 1 h, followed by an additional

2 h. at r.t. The reaction mixture was poured over crushed ice and extracted with EtOAc (4×25 mL). The organic phase was washed with aq. NH₄Cl (10%, 2×2.5 mL) and water (2×5 mL), dried with anhydrous Na₂SO₄ and concentrated to give a syrup. The crude syrupy product was purified by flash column chromatography using Hexane – EtOAc (7:3) as eluent to give corresponding epoxy sugars as colorless syrups (77–89 yields).

2.4. General procedure for ring-opening reactions of epoxy sugars with methacrylic acid

Methacrylic acid (3.35 mL, 39.4 mmol) was added to a solution of epoxy sugar in 10 mL of DMF and then TEA (0.7 mL, 5.1 mmol) was added drop-wise. The mixture was stirred for 24 h. at 65 °C and cooled by adding distilled cold water (10 mL). The resulting mixture was extracted with CH₂Cl₂, washed with water and dried over Na₂SO₄ and concentrated to give a syrup. The products were purified by flash chromatography. All the sugar-based methacrylic monomers were obtained as light yellow transparent gels (73–78% yields). [41] The resulting monomers were characterized using FT-IR, ¹H-NMR and ¹³C-NMR spectroscopies.

2.4.1. D-glufrn-MA

Colorless gel, 78% yield. $[\alpha]_D^{21}$: +0.86 (c 1, CH₂Cl₂); ¹H-NMR (400 mHz, CDCl₃): δ 6.21 (d, 1H, $J_{1,2}$: 4 Hz, H1), 6.13 (bs, 1H, H–CH=C<), 5.60 (s, 1H, H–C-CCl₃), 5.59 (d, 1H, J: 1.6 Hz, H–CH=C<), 4.92 (d, 1H, $J_{2,3}$: 0 Hz, H₂), 4.33 (dd, 1H, $J_{3,4}$: 3 Hz, $J_{4,5}$: 7.2 Hz, H₄), 3.95–4.24 (m, 7H, H₃, H₅, H_{6a}, H_{6b}, H₁₃, H_{14a}, H_{14b}), 3.50, 3.65 (dd, 1H, $J_{12a,12b}$: 10.8 Hz, $J_{12a',12b'}$: 11.2 Hz, $J_{12a,13}$: 8 Hz, $J_{12a',13}$: 3.5 Hz, H_{12a}, H_{12a'}), 3.81, 3.91 (dd, 1H, $J_{12b,13}$: 3.2 Hz, $J_{12b',13}$: 5.2 Hz, H_{12b}, H_{12b'}), 1.95 (s, 3H, methacr. CH₃), 1.36, 1.44 (2×isopr. CH₃); ¹³C-NMR (CDCl₃): δ 167.20 (>C=O), 135.91 (>C=CH₂), 126.06 (>C=CH₂), 109.71, 109.29 (2×acetal carbons), 107.02 (C₁), 99.43 (–CCl₃), 25.02, 26.82 (2×isopr. CH₃), 18.26 (methacr. CH₃).

Anal. Calcd. for $C_{18}H_{25}Cl_3O_6$: C, 43.97; H, 5.12. Found: C, 44.29; H, 5.21

2.4.2. D-manfrn-MA

White solid, 75% yield. $M_{\rm p}$ 88–90 °C; $[\alpha]_{\rm D}^{21}$ –1.16 (c 1, CH₂Cl₂); ¹H-NMR (400 mHz, CDCl₃): δ 5.92 (d, 1H, $J_{1,2}$: 4 Hz, H₁), 6.11 (bs, 1H, H–CH=C<),

5.66 (s, 1H, \underline{H} –C-CCl₃), 5.57 (d, 1H, J: 1.6 Hz, H–CH=C<), 5.06 (d, 1H, J_{2,3}: 5.6 Hz, H₂), 4.47 (m, 1H, H₅), 3.92–4.27 (m, 8H, H₃, H₄, H_{6a}, H_{6b}, H_{12b}, H₁₃, H_{14a}, H_{14b}), 3.48, 3.67 (dd, 1H, J_{12a,12b}: 10.8 Hz, J_{12a',12b'}: 11.2 Hz, J_{12a,13}: 8 Hz, J_{12a',13}: 3.5 Hz, H_{12a}, H_{12a'}), 1.93 (s, 3H, methacr. CH₃), 1.36, 1.41 (2×isopr. CH₃); ¹³C-NMR (CDCl₃): δ 167.16 (>C=O), 135.86 (>C=CH₂), 126.01 (>C=CH₂), 110.50, 110.41 (2×acetal carbons), 105.93 (C₁), 99.26 (–CCl₃), 27.09, 25.52 (2×isopr. CH₃), 18.22 (methacr. CH₃).

Anal. Calcd. for $C_{18}H_{25}Cl_3O_6$: C, 43.97; H, 5.12. Found: C, 44.02; H, 5.05

2.4.3. SD-glufrn-MA

Colorless gel, 78% yield. [α]_D²¹ –0.59 (c 1, CH₂Cl₂); ¹H-NMR (400 mHz, CDCl₃): δ 6.10 (d, 1H, $J_{1,2}$: 4 Hz, H₁), 6.12 (bs, 1H, H–CH=C<), 5.30 (s, 1H, H–C-CCl₃), 5.58 (d, 1H, J: 1.6 Hz, H–CH=C<), 4.92 (d, 1H, $J_{2,3}$: 0 Hz, H₂), 4.46 (dd, 1H, $J_{3,4}$: 3 Hz, $J_{4,5}$: 7.2 Hz, H₄), 3.94–4.34 (m, 7H, H₃, H₅, H_{6a}, H_{6b}, H₁₃, H_{14a}, H_{14b}), 3.49, 3.63 (dd, 1H, $J_{12a,12b}$: 10.8 Hz, $J_{12a',12b'}$: 11.2 Hz, $J_{12a,13}$: 8 Hz, $J_{12a',13}$: 3.5 Hz, H_{12a}, H_{12a'}), 3.79, 3.89 (dd, 1H, $J_{12b,13}$: 3.2 Hz, $J_{12b',13}$: 5.2 Hz, H_{12b}, H_{12b'}), 1.94 (s, 3H, methacr. CH₃), 1.39, 1.34 (2×isopr. CH₃); ¹³C-NMR (CDCl₃): δ 167.18 (>C=O), 135.90 (>C=CH₂), 126.00 (>C=CH₂), 109.62, 109.58 (2×acetal carbons), 106.29 (C₁), 96.83 (–CCl₃), 25.07, 26.78 (2×isopr. CH₃), 18.24 (methacr. CH₃).

Anal. Calcd. for $C_{18}H_{25}Cl_3O_6$: C, 43.97; H, 5.12. Found: C, 43.29; H, 5.09

2.4.4. D-galfrn-MA

Colorless gel, 78% yield. [α]_D²¹ –0.29 (c 1, CH₂Cl₂); ¹H NMR (400 mHz, CDCl₃): δ 6.40 (s, 1H, $\underline{\text{H}}$ –C–CCl₃), 6.19 (d, 1H, $J_{1,2}$: 4 Hz, H₁), 6.13 (bs, 1H, H–CH=C<), 5.58 (d, 1H, J: 1.6 Hz, H–CH=C<), 4.93 (d, 1H, $J_{2,3}$: 0 Hz, H₂), 3.95-4.41 (m, 8H, H₃, H₄, H₅, H_{6a}, H_{6b}, H₁₃, H_{14a}, H_{14b}), 3.56, 3.63 (dd, 1H, $J_{12a,12b}$: 10.8 Hz, $J_{12a',12b'}$: 11.2 Hz, $J_{12a,13}$: 8 Hz, $J_{12a',13}$: 3.5 Hz, H_{12a}, H_{12b}), 3.69, 3.81 (dd, 1H, $J_{12b,13}$: 3.2 Hz, $J_{12b',13}$: 5.2 Hz, H_{12a'}, H_{12b'}), 1.95 (s, 3H, methacr. CH₃), 1.37, 1.46 (2×isopr. CH₃); ¹³C-NMR (CDCl₃): δ 167.21 (>C=O), 135.93 (>C=CH₂), 126.07 (>C=CH₂), 110.25, 109.30 (2×acetal carbons), 107.10 (C₁), 99.38 (–CCl₃), 25.03, 26.42 (2×isopr. CH₃), 18.41 (methacr. CH₃).

Anal. Calcd. for $C_{18}H_{25}Cl_3O_6$: C, 43.97; H, 5.12. Found: C, 43.36; H, 5.25

2.5. General procedure for random copolymerization of sugar-based methacrylates with methyl methacrylate

Firstly, sugar based monomer (for example D-glufrn-MA, $M_w = 477.71 \text{ g} \cdot \text{mol}^{-1}$, 0.25 mmol), and methyl methacrylate (MMA, 0.4 mL, 3.75 mmol) were dissolved in deoxygenated solvent (toluene, 0.5 mL) and mixed with ligand (PMDETA, 12.5 µL, 0.06 mmol), initiator (EtBrP, 2.94 µL, 0.02 mmol) and catalysts (Cu(I)Cl, 2 mg, 0.02 mmol). And then, the mixture was degassed by three freeze-pump-thaw cycles and placed in an oil bath (90 °C) for 4 h. At the end of the reaction time by subjecting to air, the mixture was cooled to room temperature. Next, cooled mixture was diluted with THF and passed through a short silica column to remove copper salt. Then solvent was evaporated by rotary and concentrated product was precipitated in methanol, decanted and washed with methanol two times. The final product was dried under vacuum at ambient temperature for 24 h.

3. Results and discussion

Due to the recently developed controlled polymerization techniques and click chemistry reactions, synthesis of tailor-made glycopolymers has become simpler and their biological properties can be easily adjusted as a function of the type of attached carbohydrates [42–44]. For instance, a series of phenylboronic acid-based block and random glycopolymers were prepared by RAFT polymerization and their self-assembled and drug-delivery properties were systematically studied [45]. By using similar monomer compositions, the block copolymers had a more

regular transmittance change with the increasing glucose level compared to the random copolymers, however, the random copolymers exhibited a quicker insulin release rate than that of the block ones.

In this study, four different sugar-based methacrylates were firstly synthesized by two-step method: (i) etherification of protected monosaccharides 5,6-O-isopropylidene-1,2-O-(R)-trichloroethylidene-α-D-glucofuranose (**D-glufrn**), 5,6-O-isopropylidene-1,2-O-trichloroethylidene-β-D-mannofuranose (**D**manfrn), 5,6-O-isopropylidene-1,2-O-(S)-trichloroethylidene-α-D-glucofuranose (SD-glufrn), 5,6-Oisopropylidene-1,2-O-trichloroethylidene-α-D-galactofuranose (D-galfrn), with epichlorohydrin and (ii) following ring-opening reaction of obtained epoxides with metharcylic acid in the presence of triethylamine. The synthetic route and structures of obtained sugar-based monomers (D-glufrn-MA, D-manfrn-MA, SD-glufrn-MA and D-galfrn-MA) are given in Figure 1.

The chemical structures of synthesized sugar-based methacrylates were firstly characterized by FT-IR spectroscopy. As can be seen from Figure 2, the characteristic bands of monomers such as O–H, C–H, C=O, C=C, C–O–C and C–Cl bonds were clearly appeared at 3450, 2930, 1710, 1610, 1175 and 720 cm⁻¹, respectively. These results confirmed the presence of monosaccharide and methacrylate moieties in the obtained monomers.

The characteristic chemical shifts of sugar-based monomers obtained from ¹H-NMR and ¹³C-NMR spectroscopies were given in experimental part (see Chapter 2.4). The specific chemical shifts belonging

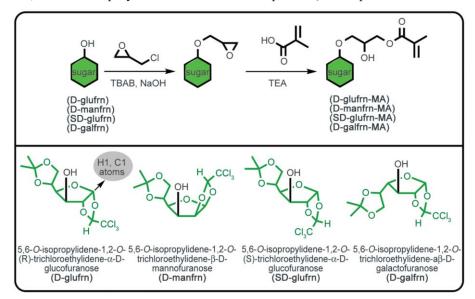


Figure 1. Synthetic pathway for the preparation of sugar based monomers

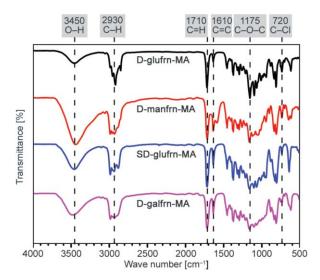


Figure 2. FT-IR spectra of D-glufrn-MA, D-manfrn-MA, SD-glufrn-MA and D-galfrn-MA

to acetal groups (>CHCCl₃) were assigned at 5.60, 5.66, 5.30 and 5.40 ppm, whereas the **H-1** protons were detected at 6.21, 5.92, 6.10 and 6.19 ppm. These shifts were in accordance with the literature data that were reported similar furanosidic trichloroethylidene acetals [46–50]. On the other hand, the characteristic protons of double bond and methyl groups of methacrylate were detected between 5.57–6.13 and 1.93–1.95 ppm, which confirmed the presence of methacrylate moieties on the monosaccharides. ¹H-NMR and ¹³C-NMR spectra of **D-glufrn-MA** were presented in Figure 3, representatively.

Additionally, the characteristic chemical shifts of C-1, CCl₃ and C(CH₃)₂ of sugar and methacrylate groups were determined by ¹³C-NMR spectroscopy and the data were given in experimental part (see Chapter 2.4). Overall, both ¹H-NMR and ¹³C-NMR results proved the chemical structures of targeted sugar-based methacrylates via etherification and ring-opening reactions. After the successful synthesis of sugar-based methacrylates, their random

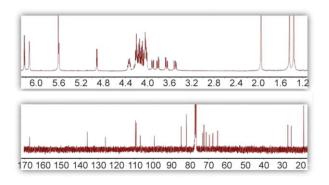


Figure 3. ¹H-NMR and ¹³C-NMR spectra of **D-glufrn-MA**

copolymerizations with methyl methacrylate (MMA) as a comonomer were investigated by classical atom transfer radical polymerization (ATRP) with a 200:1 monomer to initiator ratio. All polymerization reactions were initiated using ethyl 2-bromopropionate and complex of copper(I) chloride/N,N,N',N',N''-pentamethyldiethylenetriamine (1:1:3) at 90 °C (Figure 4).

The FT-IR spectroscopy was used to further verify the functional groups of random copolymers, where all of them displayed the similar characteristic bands of MMA and sugar-based methacrylates; a broad band at 3450 cm⁻¹ was assigned to O–H stretching vibrations, a sharp band at 1710 cm⁻¹ was related to C=O stretching vibrations, a band at 1175 cm⁻¹1 was related to C-O-C stretching vibrations, a band at 2930 cm⁻¹ was corresponded to asymmetric C-H stretching vibrations and a band at 820 cm⁻¹ was attributed to C-Cl vibrations of sugar based monomer segments (Figure 5). In addition, the successful random copolymerizations were confirmed by the absence of the C=C absorption bands at 1610 cm⁻¹ in the resulting copolymers. Overall, the chemical compositions of random copolymers containing various functional groups such as alcohol, alkyl halide ether and ester were confirmed by FT-IR analysis.

The chemical structures of random copolymers were also confirmed by ¹H-NMR analysis. As can be seen in Figure 6, the methyl (-CH₃) and methylene (-CH₂-) protons (a, c, d, e, f, h, j and u) of methacrylates were located between 0.5–2.0 ppm, whereas the characteristic methyl protons (-O-CH₃) of MMA were apparently seen at 3.62 ppm. Additionally, the chemical shifts belonging to H-1 (m) and CHCCl₃ (t) of acetal groups of sugar methacrylates were detected at 5.63–5.91 and 6.05–6.21 ppm, respectively. On the other hand, the rest protons of methylene and methine groups of sugar methacrylates were detected between 3.71 and 4.52 ppm, which confirmed the presence of methacrylate moieties on the monosaccharides. Compositions of the random copolymers could be determined from the integration ratio of methoxy protons (d at 3.62 ppm) to that of the H-1 protons (t at 5.63–5.91 ppm) [51]. The values for the composition determinations were about 94.1–97.8% and agreed well with feeding ratio (MMA:total monomer = 93.75:100). Furthermore, the molecular weight of copolymers could be also calculated from the relative intensities of the signals due to the methoxy (g at 3.62 ppm) and H-1 (t at 5.63-5.91 ppm) protons

Figure 4. Random copolymerization of sugar-based monomers with MMA by ATRP and obtained copolymers

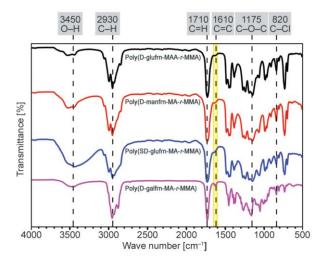


Figure 5. FT-IR spectra of poly(D-glufrn-MA-r-MMA), poly(D-manfrn-MA-r-MMA), poly(SD-glufrn-MA-r-MMA) and poly(D-galfrn-MA-r-MMA)

of methacrylate monomers divided to the CH_2 (**b** at 4.71 ppm) proton of the EtBrP initiator. According to Equation (1), the molecular weight is:

$$M_{\rm w} = \frac{\frac{g}{3}}{2b} \cdot 102.12 + \frac{t}{2b} \cdot 477.71 \tag{1}$$

where the letters represent the areas of the corresponding ¹H-NMR peaks. Overall, the molecular characteristics of synthesized copolymers were successfully evaluated by ¹H-NMR analysis.

In order to confirm the well-defined properties of random copolymers, their molecular weights and molecular weight distributions were investigated by gel permeation chromatography (GPC). As can be seen in Figure 7, all random copolymers displayed a GPC curve with monomodal and narrow size distribution. The molecular weights of the formed polymers were between 12.200 and 18.500 g/mol with relatively narrow molecular weight distributions ranging from 1.30~1.45. Furthermore, the experimental molecular weight values of copolymers obtained from the ¹H-NMR analyses were close to the theoretical values. A little differences between GPC and ¹H-NMR data were usually observed for the differences in hydrodynamic volume of copolymer's components, in which sugar-based methacrylates had considerably dissimilar structures as compared to the MMA.

The thermal properties of the obtained random copolymers were evaluated using the glass transition temperatures (T_g) determined by DSC (Table 1). Almost

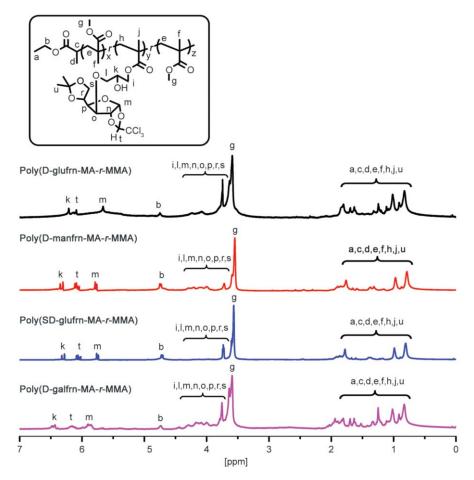


Figure 6. ¹H-NMR spectra of poly(D-glufrn-MA-*r*-MMA), poly(D-manfrn-MA-*r*-MMA), poly(SD-glufrn-MA-*r*-MMA) and poly(D-galfrn-MA-*r*-MMA)

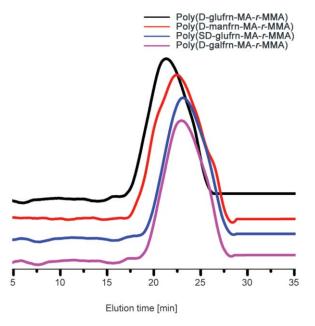


Figure 7. The GPC traces of poly(D-glufrn-MA-*r*-MMA), poly(D-manfrn-MA-*r*-MMA), poly(SD-glufrn-MA-*r*-MMA), poly(D-galfrn-MA-*r*-MMA) random copolymers via ATRP. (All measurements were conducted using tetrahydrofuran as an eluent with flow rate, 1 mL·min⁻¹ at 35 °C).

all copolymers exhibited relatively similar Tg values between 101.2 and 102.9 °C (entry 1–4). This differences could be related to either the molecular weight differences or nature of random copolymer with non-uniform composition, which was previously reported by other groups [52–54].

4. Conclusions

In conclusion, a series well-defined polymethacry-late-based random copolymers containing different methacrylates bearing protected furanosidic sugars with methyl methacrylate were successfully synthesized by ATRP. After the copolymerization, the molecular characteristics of resulting copolymers were determined by FT-IR, ¹H-NMR, GPC and DSC analyses. Their compositions were determined as 94.1~97.8%, which were agreed well with feeding ratio. On the other hand, the molecular weights, polydispersity and glass transition temperatures of the copolymers were about 10600~16800 g/mol, 1.30~1.45 and 101.2~102.9 °C respectively. Due to their structural diversity, multiple functionalities and innocuousness, the synthesis of well-defined glycopolymers is

Table 1. The random copolymerization of sugar-based methacrylates with methyl methacrylate by ATRPa

Copolymer	Conv. ^b [%]	$M_{ m n,theo}^{ m c}$ $[{ m g\cdot mol}^{-1}]$	$M_{ m n,NMR}^{ m d}$ [g·mol ⁻¹]	$M_{\mathrm{n,GPC}}^{\mathrm{e}}$ $[\mathbf{g} \cdot \mathbf{mol}^{-1}]$	Ðе	MMA ^f [%]	<i>T</i> ^g [°C]
Poly(D-glufrn-MA-r-MMA)	65.1	16400	16800	18 500	1.30	97.8	102.9
Poly(D-manfrn-MA-r-MMA)	51.6	13 100	13 200	14800	1.45	97.4	101.8
Poly(SD-glufrn-MA-r-MMA)	40.8	10400	10600	12 200	1.33	96.5	101.2
Poly(D-galfrn-MA-r-MMA)	50.4	12800	13 400	14600	1.41	95.1	102.6

^aAll polymerizations were conducted with MMA+SB-MA: EtBrP:CuCl:PMDETA: 200:1:1:3 ratio, at 90 °C for 4 h;

crucially important in many medicine and biotechnology applications.

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^bCalculated as gravimetrically;

^cThe molecular weights were calculated as follows:: $M_{n,theo} = 200/1 \cdot \text{conversion} \cdot (M_{\text{SB-MA}} + M_{\text{MMA}});$

^dThe molecular weights $(M_{n,NMR})$ were calculated using Equation (1) (from Figure 6);

^eThe molecular weight $(M_{n,GPC})$ and distribution (D) were determined by gel permeation chromatography;

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